

Effect of Dietary *Scutellaria baicalensis* Root Water Extract against *Piscicola geometra* Infection of Cobia

Putri Nurhanida Rizky^{1,2}✉, Ta-Chih Cheng¹, Happy Nursyam²

¹Department of Tropical Agriculture and International Cooperation, National Pingtung University of Science and Technology, Pingtung 91201, Taiwan

²Faculty of Fisheries and Marine Science, University of Brawijaya, Malang 65145, Indonesia

✉Corresponding's E-mail : putrimarine92@gmail.com

ABSTRACT

Piscicola geometra is an ectoparasite and the causative agent of infection in fish. Heavy infestation of *P. geometra* significantly causing mortality and serious economic losses to cobia industry. However, no single drug available today has used for the treatment or prevention of *P. geometra* infestation in fish. Water extracts of *Scutellaria baicalensis* root were evaluated for their effect on cobia infected by *P. geometra*. Fish were divided into two groups (group A for healthy fish and group B for infected fish) before being fed for 30 days with 0% as control, 0.5%, 1%, and 2% of *S. baicalensis*. Cobia growth performance, mortality, and total number of parasite infestation on fish were investigated. Result showed that infestation of *P. geometra* significantly reducing the growth performance and survival rate of cobia. Cobia fed with 1% *S. baicalensis* showed highly significant differences ($P < 0.05$) in growth performance (group A and B) and the number of parasite infestation (group B) compared to their respective controls. However, the growth performance of cobia fed with supplemented extract showed no significant differences in group A at the lowest concentration (0.5%) and the highest concentration (2%). Meanwhile group B showed significant different in growth performance among the treatment group ($P < 0.05$). This study demonstrated that *S. baicalensis* root water extract administered as a dietary supplementation is one of the most practical methods to prevent *P. geometra* infestation in cobia culture.

Original Article

PLI: S225199391700011-7

Rec. 15 Aug. 2016

Acc. 10 Sep. 2017

Pub. 25 Sep. 2017

Keywords

Oral administration, *Piscicola geometra*, *Scutellaria baicalensis*, Growth performance, Cobia, Parasite infestation

INTRODUCTION

Cobia (*Rachycentron canadum*), locally known as *badee* in Indonesia is one of the most promising candidate for marine culture industry in the world [1, 2]. The hardiness cobia is living in different environment and easy to adapt (distributed worldwide between in 50 and 1200 m depth, 16.8 to 32.2°C of temperatures, and 5 to 44.5 of salinities [3, 4]), fast growth (up to 4 – 6 kg in a year with 90% of survival rate [5]), good conversion rate (1.6 to 1) [6], and high market value (excellent white meat flesh quality) [7]. Moreover, 100 g of cobia meat contains around 32 – 507 mg of docosahexanoic acid (DHA) and 280 – 485 mg of eicosapentanoic acid (EPA) which 24% higher than Atlantic salmon that contain 10 – 11.6 mg of DHA/100g and 76 – 83 mg of EPA/100 g [8, 9]. Cobia production is expanding rapidly in Taiwan, Vietnam, China, Philippines, Indonesia, Iran, and Reunion Island [4-6, 10].

However, intensification of aquaculture production has resulted in an increased incidence of disease outbreaks as the major constraints of the mariculture industry [4]. Although there has been no detailed review of the parasites disease of cobia, the wild cobia has been reported infect by parasite at gastrointestinal tract, gills, and skin [3, 11]. Parasites infestation causing mortality, growth retardation and serious economic losses to the cobia industry [12]. *Piscicola geometra* has found to infest the eye, gills, and skin of cobia culture in Taiwan and caused a lesion, erosion, and hemorrhage. Class of Hirudinea of the Rhynchobdellida is a blood sucker leech which utilizes a proboscis to penetrate the tissue of the host [13]. The wounding makes the growth of fish slower and changes their skin color become darker, also can cause the fall of the market value [4].

Antibiotics and the veterinary drug has been applied in aquaculture to treat fish disease [14, 15]. Although the fish disease is able to be healed, the incidence of water pollution, accumulation of chemical residues, drug resistant bacteria, and highly toxic substances has become a major problem in public health [16, 17]. Nowadays, humans have returned back to nature and started to use medicinal plants as a promising alternative method to control fish disease [18, 19]. The dried root of *S. baicalensis* has been listed in Traditional Chinese Medicine (TCM) [20]. A highly active compound such as alkaloids, flavonoids, phenolics, terpenoids, steroids, pigments, and essential oils contains in *S. baicalensis* root has provided a cheaper source to treat many diseases, reducing stress responses, enhancing immune responses and mitigate many side effects with greater accuracy [21-24]. The aim of this study is to evaluate the effects of dietary *S. baicalensis* root water extract on the growth performance and disease resistance of cobia against *Piscicola geometra* infestation.

MATERIAL AND METHODS

Ethical approval

The review board and ethics committee of Tungkang Marine Biotechnology Research Station Taiwan approved the study protocol and informed consent were taken from all the participants.

Preparation of plant extract

The dried root of *S. baicalensis* was powdered mechanically using an electrical stainless grinder. The powdered (10 gram) were extracted with distilled water (150 ml) and boiled at 95°C for 20 min. The extracts were divided into four 50 ml Falcon tube and centrifuged for 10 min at 4°C, 1000 xg. The clear supernatant was stored at -20°C of refrigerator for further uses. A standard commercial diet for cobia was used throughout and designed as the control diet. Different concentration of *S. baicalensis* root water extract was sprayed into the commercial feed slowly and mixing part by part (control, 0.5%, 1%, and 2%). The diets were dried under the sterile condition in a hot air oven at 60° for 24 h.

Parasites and hosts

A total of 240 healthy cobia, with 23.3 ± 2.3 gram of body weight and 10 ± 2 cm of body length were obtained from Tungkang Marine Biotechnology Research Station, Taiwan and maintained in 200-l of tanks with air stone and water circulation. The fish were acclimatized under laboratory conditions for 7 days and fed with commercial diet for cobia. After acclimation, 120 of fish cohabited with the ones infected with *P. geometra*.

Fish and experimental protocol

The experiments were carried out at Tungkang Marine Biotechnology Research Station, Taiwan. Experimental study was divided into two group treatment. After two weeks of post-infestation, healthy fish (without leech infestation) were randomly distributed in group treatment A and infected fish in group treatment B. Each group contains four treatment including control with three replicates. Each tank (200-l) was stocked with 10 fish. Fish were fed with experimental diet in twice a day at a rate of 5% body weight for 30 days. During the experimental period, the temperature was ranged from 26 to 28°C and salinity from 33 to 35 ppt. Total number and mean body weight of fish in each tank were measured in seven days interval. The effects of treatments on the number of parasites were analyzed by comparing with those in control group after 30 days of post-experimental diet feeding.

Fish growth performance calculation

Growth performance were assessed by net weight gain (WG), specific growth rate (SGR), feed conversion ratio (FCR), and survival growth rate (SGR) at seven days interval. Calculations were made using equation was describe

[25]. The amount of the fed was increasingly based on the fish body weight. Clinical history of lesion, behavior, and mortality of the fish was observed every day. The number of leech in each fish was randomly counted in the end of experiment.

$$\text{Survival growth rate (SGR)} = \frac{\ln(\text{Final weight (g)}) - \ln(\text{Initial weight (g)}) \times 100}{\text{total duration of the experiment (days)}} \dots\dots (1)$$

$$\text{Feed Conversion Ration (FCR)} = \frac{\text{total amount of feed (g)}}{\text{weight gain (g)}} \dots\dots\dots (2)$$

$$\text{Weight gain rate (\%)} = 100 \times \frac{\text{Final body weight (g)} - \text{initial body weight (g)}}{\text{initial body weight (g)}} \dots\dots (3)$$

Statistical analysis

All data are presented as means ± standard deviation (SD). Statistical analysis was performed using SPSS 16.0 software by one-way ANOVA (analysis of variance) and accepted at the P<0.05 level. Significant differences between control and treatment groups were determined using post-hoc Duncan’s test.

RESULTS

Body weight (BW), weight gain (WG), survival growth rate (SGR), feed conversion ratio (FCR) and survival rate of cobia fed the experimental diet in both of group are shown in Tables 1 and 2. *Scutellaria baicalensis* supplemented to the fish feed were significantly enhancing the growth performance of cobia compared with control feed (P<0.05) in both of the groups. Table 1 showed that, healthy cobia fed with 1% concentration of plant extract were significantly higher than other groups treatment at the growth performance with 35.6 ± 1.2 of final body weight, 74.34 ± 5.8 of weight gain, 50.60 ± 4.1 of survival growth rate, and the FCR around 1.14 (P<0.05). According to the statistical analysis, there was no significant differences of the growth performance between the lowest concentration (0.5%) and the higher concentration (2%) in group A (P<0.05). However, no fish die in group A during the experiment.

In contrast with group A, heavy infestation of *P. geometra* caused moderate hemorrhage on skin (Figure 1) and significantly reduce the body weight of cobia after 30 days of infestation (Figure 2). Administration of 1% of *S. baicalensis* to the infected cobia slightly enhance the growth performance of fish (Figures 1 and 2) compared with control. However, there were no significant differences in weight gain of fish in group B among all treatments. There were no significant differences in survival growth rate and feed conversion ratio among fish fed with control and 0.5% or among fish fed with 1% and 2% concentration of extract (P<0.05). In addition, the survival rate in fish fed with 1% concentration of extract was significantly higher than fish fed with control, 0.5%, and 2% (P<0.05; Table 2).

Infestation of *P. geometra* in the groups treated with 1% and 2% concentration of *S. baicalensis* root were significantly reduced compared with control. There were no significant differences in parasite infestation in fish fed with commercial diet and 0.5% concentration of *S. baicalensis* root (P<0.05; Table 3).

Table 1. Weight gain, specific growth ratio, feed conversion ratio, and survival of the healthy and infected cobia fed diets containing the various concentrations of *S. baicalensis* root for 30 days.

Treatment Group A	Final body weight (g)	Weight gain ¹ (g)	SGR ² (% BW day ⁻¹)	FCR ³ (g dry feed/g gain)	Survival (%)
Control	33.6 ± 1.1 ^b	55.05 ± 5.4 ^b	39.77 ± 3.8 ^b	1.46 ± 0.08 ^b	100 ^a
0.50%	33.77 ± 1.7 ^{ab}	62.31 ± 87.2 ^{ab}	44.49 ± 5.2 ^{ab}	1.33 ± 0.08 ^{ab}	100 ^a
1%	35.6 ± 1.2 ^a	74.34 ± 5.8 ^a	50.60 ± 4.1 ^a	1.14 ± 0.06 ^a	100 ^a
2%	34.7 ± 0.7 ^{ab}	54.43 ± 4.5 ^{ab}	40.77 ± 2.6 ^{ab}	1.53 ± 0.08 ^{ab}	100 ^a
SEM	0.54	2.56	1.78	0.03	0
⁴P-value	0.037	0.02	0.045	0.011	0

Values are means from triplicate groups of fish (mean ± standard deviations n=10), where the means in each row with different superscripts are significantly different (P<0.05). ¹Weight gain= 100 x (final body weight - initial body weight) / initial body weight. ²Specific growth ratio = 100 x ln (final weight/initial weight)/ days of the experiment. ³Feed conversion ratio= g dry feed consumed/g wet weight gain. ⁴Values represent of five observations per treatment and their SEM.

Table 2. Weight gain, specific growth ratio, feed conversion ratio, and survival of infected cobia fed diets containing the various concentrations of *S. baicalensis* root for 30 days.

Treatment Group B	Final body weight (g)	Weight gain ¹ (g)	SGR ² (% BW day ⁻¹)	FCR ³ (g dry feed/g gain)	Survival (%)
Control	21 ± 0.2 ^c	21.83 ± 0.1 ^a	3.97 ± 0.1 ^b	3.25 ± 1.2 ^b	70 ^b
0.5%	21.42 ± 0 ^c	21.75 ± 1.1 ^a	1.54 ± 0.8 ^b	2.85 ± 1.1 ^b	80 ^{ab}
1%	21 ± 0.5 ^a	27.44 ± 2.3 ^a	30.69 ± 1.6 ^{ab}	1.02 ± 0.1 ^a	90 ^a
2%	22 ± 0.6 ^b	26.08 ± 5 ^a	18.56 ± 3.2 ^{ab}	1.78 ± 0.5 ^a	85 ^{ab}
SEM	0.15	1	0	1.46	3.3
⁴ P - value	0.007	0.008	0.008	0.006	0.004

Values are means from triplicate groups of fish (mean ± standard deviations n=10), where the means in each row with different superscripts are significantly different (P<0.05). ¹Weight gain = 100 x (final body weight - initial body weight) / initial body weight. ²Specific growth ratio= 100 x ln (final weight/initial weight)/ days of the experiment. ³Feed conversion ratio= g dry feed consumed/g wet weight gain. ⁴Values represent of five observations per treatment and their SEM.



Figure 1. Heavy infestation of *P. geometra* caused moderate lesion and hemorrhage on cobia skin

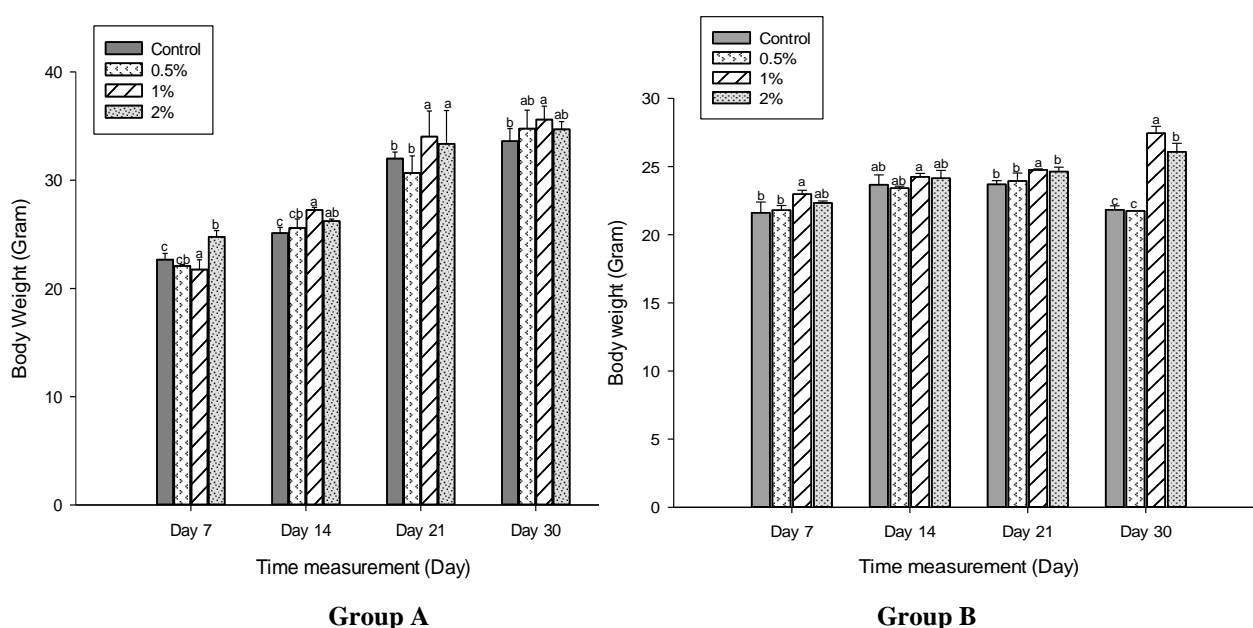


Figure 2. Body weight of healthy cobia (group A) and cobia infected by *P. geometra* (group B) fed with various concentration of *S. baicalensis* root water extract for 30 days (values are mean ± S. E). Mean value with different superscript with in a column for a parameter are significantly different (P < 0.05).

Table 3. The effectiveness of administration of *S. baicalensis* root water extract of cobia to treat *P. geometra*

Experimental group B	Number of parasites per fish for 30 days of feeding
Control	56.70 ± 7.8 ^b
0.5%	53.57 ± 9.6 ^b
1%	35.45 ± 13.5 ^a
2%	28.98 ± 18.7 ^a

Different superscripts are shows the significantly different ($P < 0.05$), intensity ± SD

DISCUSSION

Piscicola geometra is a significant cause of disease in fish, livestock and serious economic losses to the aquaculture industry. Since the used of veterinary drugs has attracted criticism, there was an urgent need to find new drugs to treat and prevent fish disease [17]. The root of *Scutellaria baicalensis* has been listed in a traditional Chinese medicine to treat human disease [24]. Although *S. baicalensis* root has been common use for human therapy for millennia, there has been tested for controlling and against fish disease [26]. These results demonstrated essentially of dietary *S. baicalensis* root for normal physiological function and had activity against parasite infestation of cobia.

In this study, the effectiveness of *S. baicalensis* root on *Piscicola geometra* was evaluated using oral administration. This study showed that heavy infestation of *P. geometra* has inhibited the growth performance of cobia. However, cobia fed with *S. baicalensis* root water extract has increased the growth performance of healthy fish (group A) and slightly improve the specific growth rate of cobia in group B (parasite infected) compared to untreated control. Administration of *S. baicalensis* also reduced the number of living parasites of 1% concentration and significantly improved the survival rate of cobia infected by *P. geometra* ($P < 0.05$). According to the Ramudu KR and Dash [27], active compounds of herbal plants supplemented on feed could induce the secretion of high protein synthesis of the digestive enzyme and stimulating the appetite and increasing food consumption and efficiencies.

The root of *S. baicalensis* produces more than 30 types of bioactive including 4'-deoxyflavones which promotes various activities such as anti-stress, growth promotion, immunostimulation, and antimicrobial activity [20]. Some study, administration of *S. baicalensis* root has significantly improved specific growth rate of olive flounder with 2% of concentration [28]. Moreover, oral administration of *S. baicalensis* root also can modulate the innate immune system of tilapia with an optimal feeding period of 3 weeks [29]. Some of the herbal extracts also have been reported to treat some parasitic disease in farm fish [26].

Water extracts of tropical seaweed *Asparagopsis taxiformis* showed the most potential for development as a natural treatment to manage the monogenean ectoparasite [30]. Dietary of garlic extract significantly reduced the infection of *Neobenedenia* sp in barramundi farmed [31]. Additionally, administrated of praziquantel resulted in over 80% reduction in worm intensity of chub mackerel culture [32]. However, freshwater bathing of praziquantel drastically increased the parasite intensity due to stress and loss of mucus during the bathing [32]. These results indicated that dietary *S. baicalensis* root could improve the growth performance and had anti-parasitic against *P. geometra* infestation with 1% of concentration.

CONCLUSION

The present study provided evidence that dietary *S. baicalensis* root could significantly enhance the growth performance and protection against parasite infection for cobia. Further work is needed to establish the stimulatory dose and optimal time of feeding of *S. baicalensis* root.

Authors' Contributions

Putri Nurhanida Rizky participated in the performed experiments, drafting the manuscript, analyze and interpretation the data. Cheng Ta-Chih participated in the design of study and financial support. Happy nursyam participated in the critically revised the manuscript for important intellectual contents. All authors of this research paper have directly participated in the planning, execution, or analysis of this study and have read and approved the final version submitted.

Competing interests

All authors declare that they have no competing interests that might have influenced the performance or presentation of the work described in this manuscript.

REFERENCES

1. Akyol O and Ünal V. 2013. Second Record of the Cobia, *Rachycentron canadum* (Actinopterygii: Perciformes: Rachycentridae), from the Mediterranean Sea. *Acta Ichthyologica et Piscatoria*. 43 (4): 315-317.
2. Gustavo A, Castellanos-Galindo, Baos R and Zapata LA. 2016. Mariculture-induced Introduction of Cobia *Rachycentron canadum* (Linnaeus, 1766), a Large Predatory Fish, in the Tropical Eastern Pacific. *BiolInvasions Records*. 5 (1): 55-58.
3. Shaffer RV and Nakamura EL. 1989. Synopsis of Biological Data on the Cobia *Rachycentron canadum* (Pisces: Rachycentridae). FAO Fisheries Synopsis 153.
4. Liao IC, Huang T-S, Tsai W-S, Hsueh C-M, Chang S-L, et al. 2004. Cobia Culture in Taiwan: Current Status and Problems. *Aquaculture*. 237 (1-4): 155-165.
5. Benetti DD, O'Hanlon B, Rivera JA, Welch AW, Maxey C, et al. 2010. Growth Rates of Cobia (*Rachycentron canadum*) Cultured in Open Ocean Submerged Cages in the Caribbean. *Aquaculture*. 302 (3-4): 195-201.
6. Nhu VC, Nguyen HQ, Le TL, Tran MT, Sorgeloos P, et al. 2011. Cobia *Rachycentron canadum* Aquaculture in Vietnam: Recent Developments and Prospects. *Aquaculture*. 315 (1-2): 20-25.
7. Moreira CB, Rombenso AN, Candioto FB and Tsuzuki MY. 2015. Feeding Frequency Affects Growth of Juvenile Cobia *Rachycentron canadum* Cultured in Near-shore Cages. *Bol. Inst. Pesca, São Paulo*. 41 (2): 219-226.
8. Allison EH. 2011. Aquaculture, fisheries, poverty and food security. Secondary title.
9. Guo JJ, Kuo CM, Hong JW, Chou RL, Lee YH, et al. 2015. The Effects of Garlic-supplemented Diets on Antibacterial Activities Against *Photobacterium damsela* subsp. *Piscicida* and *Streptococcus iniae* and on Growth in Cobia, *Rachycentron canadum*. *Aquaculture*. 435: 111-115.
10. Musthofa SZ, Zamroni M and Kadarini T. 2016. Penggunaan Probiotik Komersial pada Pemeliharaan Larva Ikan Rainbow Kurumoi (*Melanotaenia parva*). Secondary title; p. 901-907.
11. Cnaani A, McLean E and Hallerman EM. 2013. Effects of Growth Hormone Transgene Expression and Triploidy on Acute Stress Indicators in Atlantic Salmon (*Salmo salar* L.). *Aquaculture*. 412-413: 107-116.
12. Chu KB, Abdulah A, Abdullah SZ and Bakar RA. 2013. A Case Study on the Mortality of Cobia (*Rachycentron canadum*) Cultured in Traditional Cages. *Tropical Life Sciences Research*. 24 (2): 77-84.
13. Mann KH. 1962. Chapter 2 - The Medicinal Leech, *hirudo medicinalis*. Book title: Pergamon; p. 5-21.
14. Lee C-S. 2015. Dietary nutrients, Additives and Fish health: John Wiley & Sons. ISBN: 9781119005537.
15. Rico A, Phu TM, Satapornvanit K, Min J, Shahabuddin AM, et al. 2013. Use of veterinary Medicines, Feed Additives and Probiotics in Four Major Internationally Traded Aquaculture Species Farmed in Asia. *Aquaculture*. 412-413: 231-243.
16. Reverter M, Bontemps N, Lecchini D, Banaigs B and Sasal P. 2014. Use of Plant Extracts in Fish Aquaculture as an Alternative to Chemotherapy: Current Status and Future Perspectives. *Aquaculture*. 433: 50-61.
17. Rico A and Van den Brink PJ. 2014. Probabilistic Risk Assessment of Veterinary Medicines Applied to Four Major Aquaculture Species Produced in Asia. *Science of the Total Environment*. 468: 630-641.
18. Rafieian-Kopaei M. 2013. Medicinal Plants and the Human Needs. *Journal of HerbMed Pharmacology*. 1 (1): 1-2.
19. Syahidah A, Saad C, Daud H and Abdelhadi Y. 2015. Status and Potential of Herbal Applications in Aquaculture: A. *Iranian J Fish Sci*. 14 (1): 27-44.
20. Zhao Q, Chen X-Y and Martin C. 2016. *Scutellaria baicalensis*, the Golden Herb from the Garden of Chinese Medicinal Plants. *Sci Bull*. 61 (18): 1391-1398.
21. Shirzad H and Nasri H. 2014. Toxicity and Safety of Medicinal Plants. *J HerbMed Pharmacol*. 2 (2): 21-22.
22. Van Hai N. 2015. The Use of Medicinal Plants as Immunostimulants in Aquaculture: A review. *Aquaculture*. 446: 88-96.
23. Zhu D, Wang S, Lawless J, He J and Zheng Z. 2016. Dose Dependent Dual Effect of Baicalin and Herb Huang Qin Extract on Angiogenesis. *PLoS ONE*. 11 (11): e0167125.
24. Muluye RA, Bian Y and Alemu PN. 2014. Anti-inflammatory and Antimicrobial Effects of Heat-clearing Chinese Herbs: A Current Review. *J Tradition Complemen Med*. 4 (2): 93-98
25. Strand Å. 2005. Growth and Bioenergetic Models and Their Application in Aquaculture of Perch (*Perca fluviatilis*): Vattenbruksinstitutionen, Sveriges lantbruksuniversitet. ISSN 1101-6620.
26. Harikrishnan R, Balasundaram C and Heo M-S. 2011. Impact of Plant Products on Innate and Adaptive Immune System of Cultured Finfish and Shellfish. *Aquaculture*. 317 (1-4): 1-15.
27. Ramudu KR and Dash G. 2013. A Review on Herbal Drugs Against Harmful Pathogens in Aquaculture. *American J Drug Disc Develop*. 3 (4): 209-219.

28. Cho SH, Jeon GH, Kim HS, Kim DS and Kim C. 2013. Effects of Dietary *Scutellaria baicalensis* Extract on Growth, Feed Utilization and Challenge Test of Olive Flounder (*Paralichthys olivaceus*). *Asian-Australasian J Animal Sci.* 26 (1): 90-96.
29. Yin G, Jeney G, Racz T, Xu P, Jun X, et al. 2006. Effect of Two Chinese Herbs (*Astragalus radix* and *Scutellaria radix*) on Non-Specific Immune Response of Tilapia, *Oreochromis niloticus*. *Aquaculture.* 253 (1–4): 39-47.
30. Hutson KS, Mata L, Paul NA and de Nys R. 2012. Seaweed Extracts as a Natural Control Against the Monogenean Ectoparasite, *Neobenedenia* sp., Infecting Farmed Barramundi (*Lates calcarifer*). *Intern J Parasitol.* 42 (13): 1135-1141.
31. Militz TA, Southgate PC, Carton AG and Hutson KS. 2013. Dietary Supplementation of Garlic (*Allium sativum*) to Prevent Monogenean Infection in Aquaculture. *Aquaculture.* 408: 95-99.
32. Yamamoto S, Shirakashi S, Morimoto S, Ishimaru K and Murata O. 2011. Efficacy of Oral Praziquantel Treatment Against the Skin Fluke Infection of Cultured Chub Mackerel, *Scomber japonicus*. *Aquaculture.* 319 (1): 53-57.